

Communication

## Colonies of Bumble Bees (*Bombus impatiens*) Produce Fewer Workers, Less Bee Biomass, and Have Smaller Mother Queens Following Fungicide Exposure

Olivia M. Bernauer <sup>1,†</sup>, Hannah R. Gaines-Day <sup>1,†</sup> and Shawn A. Steffan <sup>1,2,\*</sup>

<sup>1</sup> Department of Entomology, University of Wisconsin, 1630 Linden Dr., Madison, WI 53706, USA; E-Mails: ombernauer@wisc.edu (O.M.B.); hgaines@wisc.edu (H.R.G.-D.)

<sup>2</sup> United States Department of Agriculture, Agricultural Research Service, Vegetable Crops Research Unit, Madison, WI 53706, USA

† These authors contributed equally to this work.

\* Author to whom correspondence should be addressed; E-Mail: steffan@entomology.wisc.edu; Tel.: +1-608-890-1281; Fax: +1-608-262-3322.

Academic Editor: Brian T. Forschler

Received: 18 March 2015 / Accepted: 27 May 2015 / Published: 1 June 2015

---

**Abstract:** Bees provide vital pollination services to the majority of flowering plants in both natural and agricultural systems. Unfortunately, both native and managed bee populations are experiencing declines, threatening the persistence of these plants and crops. Agricultural chemicals are one possible culprit contributing to bee declines. Even fungicides, generally considered safe for bees, have been shown to disrupt honey bee development and impair bumble bee behavior. Little is known, however, how fungicides may affect bumble bee colony growth. We conducted a controlled cage study to determine the effects of fungicide exposure on colonies of a native bumble bee species (*Bombus impatiens*). Colonies of *B. impatiens* were exposed to flowers treated with field-relevant levels of the fungicide chlorothalonil over the course of one month. Colony success was assessed by the number and biomass of larvae, pupae, and adult bumble bees. Bumble bee colonies exposed to fungicide produced fewer workers, lower total bee biomass, and had lighter mother queens than control colonies. Our results suggest that fungicides negatively affect the colony success of a native bumble bee species and that the use of fungicides during bloom has the potential to severely impact the success of native bumble bee populations foraging in agroecosystems.

**Keywords:** chlorothalonil; bee decline; native bees

---

## 1. Introduction

Bees are among the most important pollinators, providing vital services to natural and agricultural systems. Approximately 90% of flowering plants, including 35% of all crops, require insect mediated pollination [1,2]. Managed honey bees are the most widely used bees for crop pollination, although for certain crops, native bees are more effective pollinators than honey bees (e.g., cranberry, [3]; apple, [4]; cherry, [5]; alfalfa, [6]). Unfortunately, both managed honey bees and wild, native bees have been declining around the world [7–10]. Although the exact cause of these declines is uncertain, several factors have been implicated including habitat loss and fragmentation, disease, and exposure to agricultural chemicals [7,11,12]. Gaining a better understanding of each of the factors driving these declines is vital to ensure the continued provisioning of pollination services to both natural and agricultural systems.

Exposure to agricultural chemicals is one factor implicated in bee declines that has received much attention in both the scientific and popular media [13–18]. The majority of research focuses on insecticides (e.g., [19–21]); however, increasing evidence suggests that fungicides may also have detrimental effects on bees [22–25]. For example, honey bee colonies exposed to chlorothalonil, a commonly used, broad-spectrum fungicide, had higher rates of mortality than colonies not exposed to the chemical [24]. Furthermore, honey bee larvae reared on pollen contaminated with fungicides (Captan<sup>®</sup>, Rovral<sup>®</sup>, or Ziram<sup>®</sup>) failed to develop into adults [25]. Additionally, Pettis *et al.* [26] found that 100% of honey bee-collected pollen in agricultural landscapes contained fungicide residue. This suggests that fungicide exposure to honey bees is common and frequent. Since wild, native bees are also important crop pollinators, it is likely that they, too, run the risk of being exposed to fungicides if foraging within agricultural landscapes. It is critical, therefore, to examine the effects of fungicides on both managed and native bees.

Bumble bees are known to be important pollinators in many systems [1,27,28] and are unique among bees native to North America because they exist both as wild colonies as well as commercially available managed colonies, making them a model organism to study how agrochemical exposure affects wild bees. Of the few studies that have investigated the effect of fungicides on bumble bees, the results were conflicting (e.g., altered foraging behavior, [23], no effect [29,30] no effect on brood production yet repellency to adults [31]). Few studies, therefore, have documented the effect of fungicides on bumble bee colony growth and success. Here, we have conducted a controlled caging study to isolate the impacts of the fungicide, chlorothalonil, on the colony success of a North American bumble bee species (*Bombus impatiens* Cresson). Chlorothalonil is a widely used fungicide on crop and ornamental plants [32] and is commonly found in high concentrations within the pollen and hives of honey bees foraging in agricultural systems [14,26]. Well-known products that contain chlorothalonil as their active ingredient include Bravo<sup>®</sup>, Daconil<sup>®</sup>, and Sweep<sup>®</sup> [33]. The initial intended use of chlorothalonil was for fungal diseases of turf grasses [34], but it was later approved for use on food crops and is now applied to a wide variety of flowering nut, vegetable, and fruit crops [33]. The application of chlorothalonil to flowering agricultural crops means that there is the potential for pollinators to come in contact with this

compound. For example, Mullin *et al.* [14] surveyed honey bee colonies from 23 states and found that chlorothalonil was the most frequently detected pesticide in pollen and wax samples. Additionally, chlorothalonil has been found within “entombed pollen” in honey bee colonies that suffered from colony collapse disorder (CCD) [24]. The fact that honey bees are commonly exposed to chlorothalonil suggests that bumble bees and other native species foraging in agricultural crops are also being exposed. Given the negative impacts associated with chlorothalonil exposure in honey bees [24], we hypothesize that bumble bee colonies exposed to this fungicide will have reduced colony success, as measured by number and biomass of larvae, pupae, and adult bees.

## 2. Materials and Methods

### 2.1. Experimental Set-Up

To assess the effect of fungicide exposure on bumble bee colony success, we conducted a cage study at the West Madison Agricultural Research Station (Madison, WI). Ten mesh cages (3 m × 3 m base × 2 m tall, Ozark Trail Instant 10' × 10' Screen Houses) were set up in a field planted with oats (*Avena barbata*). A trench was dug around each cage and all four edges were buried into the ground to ensure that bees could not escape. Cages were stocked with potted, flowering plants known to be attractive to bees: buckwheat (*Fagopyrum esculentum*), borage (*Borago officinalis*), alyssum (*Lobularia maritima* cultivar), cosmos (*Cosmos bipinnatus* cultivars), and sunflowers (*Helianthus annuus* cultivars). Cages were additionally supplemented with a single tray (36 × 42 cm) of in-bloom clover (*Trifolium* sp.). Floral resources were clustered within one corner of the cage occupying a space approximately 2.5 m × 1 m. The remaining area of the cage was evenly vegetated by the oats. Bumble bee colonies (*Bombus impatiens*, research “mini-colonies,” Koppert Biological Systems, Howell, MI) each containing workers and a single queen, were randomly assigned a treatment (fungicide +/-,  $n = 5$  colonies per treatment), then placed within a field cage ( $n = 1$  colony per cage) for 29 days (23 June–21 July 2014). Initial colony weight did not differ between treatments ( $F_{1,9} = 0.18$ ,  $p = 0.68$ ). Colonies were retrieved from the field prior to producing gynes and placed in a freezer.

To protect the colony boxes from direct sun and rain, colonies were placed in plastic crates wrapped in insulated bubble wrap and set on two bricks. Crates were staked down with two large turf stakes and a piece of parachute cord. Colony boxes were oriented so that the colony’s opening was facing south to provide the bees with optimal navigational conditions [35]. Colonies were provided sugar water bladders to supplement nectar availability (similar to [17]), which remained in place for the duration of the study.

The fungicide chlorothalonil (Bravo<sup>®</sup> with Weatherstik, Syngenta AG, Basel, Switzerland) was applied at a field-relevant level (20 g/L) to flowering plants in the five fungicide treatment cages using a hand held pesticide sprayer (RL FloMaster Yard and Garden Sprayer, 1 Gallon, Model 1401P) twice during the study (day 0 and 13). Fungicide was applied as “spray to drip,” meaning that all flowers were uniformly coated such that no further liquid could adhere to floral surfaces. Approximately 0.8 L of liquid was applied per cage. This corresponds to ~17 kg/ha of the active ingredient, chlorothalonil, which is below the upper range of allowable rates for ornamental plants and turf. Fungicide was only applied to plants. Colonies were present in the cages when the sprays occurred, but colonies were not sprayed directly. Applications were made in the late evening at and after dusk and on days with little or no wind or rain.

## 2.2. Sampling and Analyses

At the conclusion of the study, *B. impatiens* colonies were collected from the cages. Exit doors were blocked 24 h prior to collection, allowing bees to return to the colony box but not leave. Any bees that were seen outside the colony at the time of retrieval were collected into a vial and kept with their respective colony. The following response variables were measured: number and dry-weight of larvae, pupae, workers (*i.e.*, adult female foragers), adult males, and the mother queen.

Before analyzing the data, we used a Shapiro-Wilk Goodness of Fit test to assess whether the data met the assumptions of normality and 2-sided *F*-test to assess whether variances were equal. For variables that met the assumptions, a one-way ANOVA was performed with treatment as the independent variable. For data that did not meet the assumptions, a non-parametric Wilcoxon Rank Sum Test (also called Mann-Whitney *U* test) was performed. Statistics related to normality and variance are presented in Table 1. All statistical analyses were conducted in JMP Pro 11 [36].

## 3. Results

Colonies exposed to the fungicide treatment ( $n = 5$ ) produced less than a third as many workers (mean  $\pm$  SE, fungicide  $12.2 \pm 3.8$  vs. control  $43.2 \pm 11.2$ ,  $F_{1,9} = 6.8$ ,  $p = 0.03$ ), less than half the amount of bee biomass (fungicide  $0.91\text{g} \pm 0.15$  vs. control  $2.36\text{g} \pm 0.55$ ,  $F_{1,9} = 8.3$ ,  $p = 0.02$ ) and had mother queens with half the body mass (fungicide  $0.14\text{g} \pm 0.04$  vs. control  $0.27 \pm 0.01$ ,  $Z = 2.5$ ,  $p = 0.01$ ) as compared to colonies in the no-fungicide control treatment (Table 1, Figure 1). The number of larvae, pupae, and males did not differ between treatments (Table 1). The biomass of each individual life stage (larvae, pupae, workers, and adult males) also did not differ (Table 1), nor did the weight per individual for larvae, pupae, and workers (Table 1). There were insufficient data to analyze the weight per individual for adult males by treatment.

## 4. Discussion

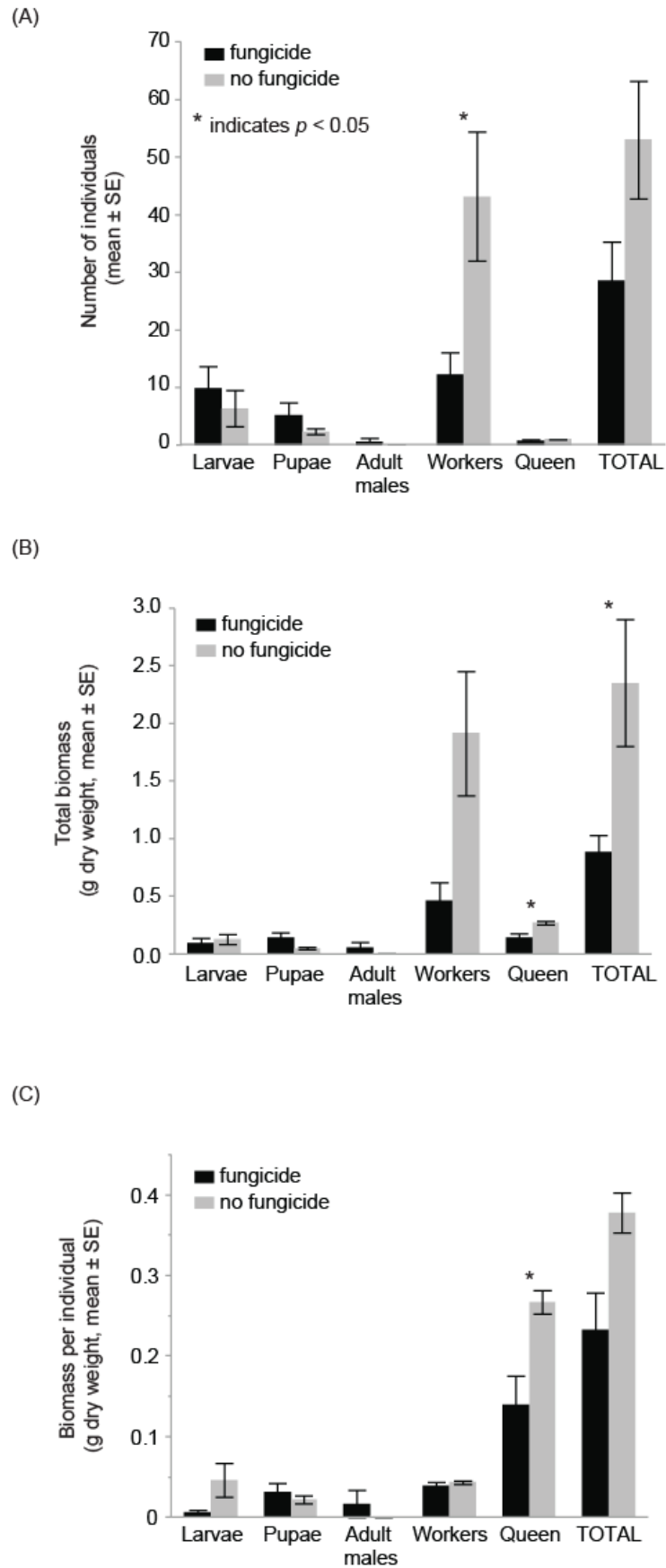
Over the past decade, declines among certain bee taxa have been documented worldwide [11]. One possible factor contributing to their decline is exposure to agricultural chemicals. Fungicides, used widely in agricultural systems, are generally considered safe for bees [29,31,37], but recent findings suggest otherwise (e.g., [22,23,25,30]). Using field cages, we found that bumble bee colonies exposed to the fungicide, chlorothalonil, resulted in significantly fewer worker bees, less total bee biomass and smaller mother queens than those in the control treatment.

One of the clearest patterns in our study was the significant reduction in the number of worker bees within the colonies exposed to fungicide. Colony success is dependent on the capacity of workers to collect resources and care for the queen, allowing the colony to grow and ultimately produce as many new queens as possible [38]. In order to maximize queen production, the colony must first collect sufficient resources to maximize forager numbers. With fewer foragers, a colony will be compromised in its capacity to support new queens. Thus, a colony that has a reduced number of foragers will likely have reduced fitness [38,39].

**Table 1.** Results of statistical analysis comparing the number, biomass, and biomass per individual for larvae, pupae, and adult (*i.e.*, female, male, and mother queen) bumble bees in fungicide treatments as compared to the non-fungicide control. For data that did not meet the assumptions of normality or equal variance, a non-parametric test was used.

| Y-variable             |             | Shapiro-Wilk †             | F-test, 2-sided ‡               | ANOVA                       | Wilcoxon Rank Sum     |
|------------------------|-------------|----------------------------|---------------------------------|-----------------------------|-----------------------|
| Number                 | Larvae      | $W = 0.92, p = 0.37$       | $F_{4,4} = 1.5, p = 0.71$       | $F_{1,9} = 0.47, p = 0.51$  |                       |
|                        | Pupae       | $W = 0.96, p = 0.84$       | $F_{4,4} = 17.8, p = 0.02 *$    |                             | $Z = -0.74, p = 0.46$ |
|                        | Workers     | $W = 0.97, p = 0.92$       | $F_{4,4} = 8.5, p = 0.06$       | $F_{1,9} = 6.8, p = 0.03 *$ |                       |
|                        | Adult males | $W = 0.62, p = 0.0001 ***$ | insufficient data               |                             | $Z = -0.8, p = 0.42$  |
|                        | Total       | $W = 0.96, p = 0.80$       | $F_{4,4} = 2.3, p = 0.43$       | $F_{1,9} = 4.0, p = 0.08$   |                       |
| Biomass                | Larvae      | $W = 0.95, p = 0.67$       | $F_{4,4} = 1.1, p = 0.93$       | $F_{1,9} = 0.27, p = 0.62$  |                       |
|                        | Pupae       | $W = 0.89, p = 0.17$       | $F_{4,4} = 13.1, p = 0.03 *$    |                             | $Z = -1.5, p = 0.14$  |
|                        | Workers     | $W = 0.94, p = 0.58$       | $F_{4,4} = 12.1, p = 0.03 *$    |                             | $Z = 1.7, p = 0.09$   |
|                        | Adult males | $W = 0.62, p = 0.0001 ***$ | insufficient data               |                             | $Z = -0.8, p = 0.42$  |
|                        | √(total)    | $W = 0.96, p = 0.79$       | $F_{4,4} = 5.6, p = 0.12$       | $F_{1,9} = 8.3, p = 0.02*$  |                       |
| Biomass per individual | Larva       | $W = 0.85, p = 0.08$       | $F_{4,4} = 101.5, p = 0.003 **$ |                             | $Z = -1.1, p = 0.62$  |
|                        | Pupa        | $W = 0.94, p = 0.63$       | $F_{3,4} = 3.9, p = 0.22$       | $F_{1,8} = 2.2, p = 0.18$   |                       |
|                        | Workers     | $W = 0.96, p = 0.78$       | $F_{4,4} = 5.1, p = 0.14$       | $F_{1,9} = 0.80, p = 0.40$  |                       |
|                        | Adult male  | insufficient data          | insufficient data               |                             | insufficient data     |
|                        | Queen       | $W = 0.81, p = 0.02 *$     | $F_{4,4} = 6.2, p = 0.11$       |                             | $Z = 2.5, p = 0.01*$  |

†  $p < 0.05$  indicates that residuals are not normally distributed and transformation or use of non-parametric test is required; ‡  $p < 0.05$  indicates that variances are not the same and transformation or use of non-parametric test is required; \* indicates  $p < 0.05$ ; \*\* indicates  $p < 0.01$ ; \*\*\* indicates  $p < 0.001$ .



**Figure 1.** Bumble bee colonies exposed to the fungicide chlorothalonil had (A) fewer workers, (B) lower total bee biomass and (C) smaller mother queens than control colonies. Statistics for each life stage comparison can be found in Table 1.

Another pattern observed in our study was the significantly lower mother queen weight in colonies exposed to fungicides compared to the no-fungicide controls. Since the fungicide treated colonies also had fewer foragers, it is likely that the mother queens were not receiving sufficient food. Furthermore, a previous study has shown that fungicide exposure results in impaired bee foraging efficiency [23] suggesting that ostensibly unaffected workers may not be collecting adequate resources.

It should be noted that these patterns were observed within large field cages, an unnatural setting for a wild bee species. While this manipulation was critical to confine bee foraging to flowers with fungicide residue, we acknowledge that it likely introduced a degree of contrivance, and thus our results should be viewed in light of this. It should also be noted that any cage-effect was equally applicable to the colonies in the control cages, which were subjected to the same biotic and abiotic circumstances. Another limitation of our study was our relatively small sample sizes (five replications each of the fungicide/no-fungicide treatments). Greater sample sizes are often helpful to distinguish among treatments, particularly where variances are large, but our sample sizes provided adequate data to discern treatment effects. Another element of our study that might have been improved was the inclusion of bee-counts and/or measurement of total bee biomass prior to the start of the experiment. We did measure total colony biomass prior to the start of the study, but we did not specifically weigh or count the bees in each colony. Doing so may have compromised bee health to an unknown degree, so we opted to weigh the entire colony instead. No significant differences were found among any of the founding colonies. Finally, at the conclusion of our study we measured multiple response variables relevant to bee fitness but could not measure queen production because by mid-season, the colonies had not yet begun to produce new queens. However, measurement of mid-season forager abundance represents a closely linked proxy for queen productivity, given that colonies with more foragers are known to produce more queens [38,39].

While previous research suggests that fungicides are not lethal to adult bees [29,31,37], our results demonstrate that fungicide exposure results in smaller colony size in the native North American bumble bee, *Bombus impatiens*. These results suggest that a mechanism other than acute toxicity is responsible for lower colony success. There are many potential factors that may be driving these impacts on *B. impatiens*. One possible mechanism is a disruption of the microbial community associated with bees. Previous researchers have documented strong associations between bees and microbes, including bacteria and fungi, especially yeasts [40,41]. This research suggests that microbes play an important role in the health of bees by, for example, preventing pollen provisions from spoiling and providing added vitamins (e.g., yeasts produce critical B vitamins, [42]) to their diet. If pollen collected by bees is contaminated with fungicides, it stands to reason that the community of beneficial microorganisms, specifically fungi, may be altered, and the digestive and nutritional benefits provided by these microbes could be compromised. One study has already demonstrated changes in the fungal community in pollen collected by honey bees after exposure to fungicides [43] suggesting that similar patterns may be true for other bees. Future research should characterize how fungicide residues may be altering the microbiome of pollen-provisions, and thus the nutrition of native bees.

## 5. Conclusions

We provide empirical evidence that one particular fungicide, chlorothalonil, can cause significant colony losses in bumble bees. Specifically, we report that the number of worker bees in *Bombus impatiens*

colonies was reduced from a mean of 43.2 bees in control cages to 12.2 in fungicide-treated cages. Such losses drove the significant reduction in total colony biomass, a consequence that directly affects the production of new queens in the fall [38,39]. Additionally, the mean weight of mother queens among the fungicide-treated colonies was significantly reduced from that of controls, indicating there were sub-lethal consequences of chlorothalonil. These findings call into question whether fungicides are completely bee-safe for all bee species, and point to the need for further studies examining the acute and chronic impacts of fungicides on native bees. The trade-offs associated with fungicide use during bloom will also likely need to be re-visited.

### Acknowledgments

The authors would like to thank the following people for their assistance in the field and lab: Janet van Zoeren, Elissa Chasen, Chris Watson, Scott Lee, Annie Deutsch. We would also like to thank the staff at the West Madison Agricultural Research Station. This work was supported by the University of Wisconsin Vilas Lifecycle Professorship (awarded to S.A.S.) and the USDA-Agricultural Research Service appropriated funds (Current Research Information System #3655-21220-001).

### Author Contributions

Hannah R. Gaines-Day, Shawn A. Steffan, and Olivia M. Bernauer conceived and designed the experiments; Olivia M. Bernauer, Hannah R. Gaines-Day, and Shawn A. Steffan performed the experiments; Hannah R. Gaines-Day and Shawn A. Steffan analyzed the data; Hannah R. Gaines-Day, Olivia M. Bernauer, and Shawn A. Steffan wrote the paper.

### Conflicts of Interest

The authors declare no conflict of interest.

The founding sponsors had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, and in the decision to publish the results.

### References

1. Klein, A.-M.; Vaissière, B.E.; Cane, J.H.; Steffan-Dewenter, I.; Cunningham, S.A.; Kremen, C.; Tscharntke, T. Importance of pollinators in changing landscapes for world crops. *Proc. R. Soc. B Biol. Sci.* **2007**, *274*, 303–313.
2. Ollerton, J.; Winfree, R.; Tarrant, S. How many flowering plants are pollinated by animals? *Oikos* **2011**, *120*, 321–326.
3. Cane, J.H.; Schiffhauer, D. Dose-response relationships between pollination and fruiting refine pollinator comparisons for cranberry (*Vaccinium macrocarpon* [Ericaceae]). *Am. J. Bot.* **2003**, *90*, 1425–1432.
4. Vicens, N.; Bosch, J. Pollinating efficacy of *Osmia cornuta* and *Apis mellifera* (Hymenoptera: Megachilidae, Apidae) on “red Delicious” apple. *Environ. Entomol.* **2000**, *29*, 235–240.
5. Bosch, J.; Kemp, W.P.; Trostle, G.E. Bee population returns and cherry yields in an orchard pollinated with *Osmia lignaria* (Hymenoptera: Megachilidae). *J. Econ. Entomol.* **2006**, *99*, 408–413.



6. Pitts-Singer, T.L.; Cane, J.H. The Alfalfa Leafcutting Bee, *Megachile rotundata*: The World's Most Intensively Managed Solitary Bee. *Annu. Rev. Entomol.* **2011**, *56*, 221–237.
7. Potts, S.G.; Biesmeijer, J.C.; Kremen, C.; Neumann, P.; Schweiger, O.; Kunin, W.E. Global pollinator declines: Trends, impacts and drivers. *Trends Ecol. Evol.* **2010**, *25*, 345–353.
8. Ellis, J.D.; Evans, J.D.; Pettis, J. Colony losses, managed colony population decline, and Colony Collapse Disorder in the United States. *J. Apic. Res.* **2010**, *49*, 134–136.
9. Bartomeus, I.; Ascher, J.S.; Gibbs, J.; Danforth, B.N.; Wagner, D.L.; Hedtke, S.M.; Winfree, R. Historical changes in northeastern US bee pollinators related to shared ecological traits. *Proc. Natl. Acad. Sci.* **2013**, *110*, 4656–4660.
10. Goulson, D.; Lye, G.C.; Darvill, B. Decline and conservation of bumble bees. In *Annual Review of Entomology*; Annual Reviews: Palo Alto, CA, USA, 2008; Volume 53, pp. 191–208.
11. Goulson, D.; Nicholls, E.; Botías, C.; Rotheray, E.L. Bee declines driven by combined stress from parasites, pesticides, and lack of flowers. *Science* **2015**, *347*, doi:10.1126/science.1255957.
12. Gonzalez-Varo, J.P.; Biesmeijer, J.C.; Bommarco, R.; Potts, S.G.; Schweiger, O.; Smith, H.G.; Steffan-Dewenter, I.; Szentgyörgyi, H.; Woyciechowski, M.; Vila, M. Combined effects of global change pressures on animal-mediated pollination. *Trends Ecol. Evol.* **2013**, *28*, 524–530.
13. Johnson, R.M.; Ellis, M.D.; Mullin, C.A.; Frazier, M. Pesticides and honey bee toxicity—USA. *Apidologie* **2010**, *41*, 312–331.
14. Mullin, C.A.; Frazier, M.; Frazier, J.L.; Ashcraft, S.; Simonds, R.; vanEngelsdorp, D.; Pettis, J.S. High levels of miticides and agrochemicals in North American apiaries: Implications for honey bee health. *PLoS ONE* **2010**, *5*, e9754.
15. Zhu, W.; Schmehl, D.R.; Mullin, C.A.; Frazier, J.L. Four common pesticides, their mixtures and a formulation solvent in the hive environment have high oral toxicity to honey bee larvae. *PLoS ONE* **2014**, *9*, e77547.
16. Johnson, R.M.; Dahlgren, L.; Siegfried, B.D.; Ellis, M.D. Acaricide, fungicide and drug interactions in honey bees (*Apis mellifera*). *PLoS ONE* **2013**, *8*, e54092.
17. Morandin, L.A.; Winston, M.L.; Franklin, M.T.; Abbott, V.A. Lethal and sub-lethal effects of spinosad on bumble bees (*Bombus impatiens* Cresson). *Pest Manag. Sci.* **2005**, *61*, 619–626.
18. Beeing there: The search for pesticides' effect on declining bee colonies moves to the fields. Available online: <http://www.scientificamerican.com/article/beeing-there-the-search-for-pesticides-effect-on-declining-bee-colonies-moves-to-the-fields/> (accessed on 17 March 2015).
19. Sterk, G.; Heuts, F.; Merck, N.; Bock, J. Sensitivity of non-target arthropods and beneficial fungal species to chemical and biological plant protection products: Results of laboratory and semi-field trials. In Proceedings of the International Symposium on Biologocial Control of Arthropods, Pucón, Chile, 4–8 March 2013.
20. Whitehorn, P.R.; O'Connor, S.; Wackers, F.L.; Goulson, D. Neonicotinoid pesticide reduces bumble bee colony growth and queen production. *Science* **2012**, *336*, 351–352.
21. Barbosa, W.F.; de Meyer, L.; Guedes, R.N.C.; Smagghe, G. Lethal and sublethal effects of azadirachtin on the bumblebee *Bombus terrestris* (Hymenoptera: Apidae). *Ecotoxicology* **2012**, *24*, 130–142.
22. Ladurner, E.; Bosch, J.; Kemp, W.P.; Maini, S. Assassing delayed and acute toxicity of five formulated fungicides to *Osmia lignaria* Say and *Apis mellifera*. *Apidologie* **2005**, *36*, 449–460.

23. Sprayberry, J.D.H.; Ritter, K.A.; Riffell, J.A. The Effect of olfactory exposure to non-insecticidal agrochemicals on bumblebee foraging behavior. *PLoS ONE* **2013**, *8*, e76273.
24. vanEngelsdorp, D.; Evans, J.D.; Donovall, L.; Mullin, C.; Frazier, M.; Frazier, J.; Tarpy, D.R.; Hayes, J., Jr.; Pettis, J.S. “Entombed Pollen”: A new condition in honey bee colonies associated with increased risk of colony mortality. *J. Invertebr. Pathol.* **2009**, *101*, 147–149.
25. Mussen, E.C.; Lopez, J.E.; Peng, C.Y.S. Effects of selected fungicides on growth and development of larval honey bees, *Apis mellifera* L. (Hymenoptera: Apidae). *Environ. Entomol.* **2004**, *33*, 1151–1154.
26. Pettis, J.S.; Lichtenberg, E.M.; Andree, M.; Stitzinger, J.; Rose, R.; vanEngelsdorp, D. Crop pollination exposes honey bees to pesticides which alters their susceptibility to the gut pathogen *Nosema ceranae*. *PLoS ONE* **2013**, *8*, e70182.
27. Greenleaf, S.S.; Kremen, C. Wild bee species increase tomato production and respond differently to surrounding land use in Northern California. *Biol. Conserv.* **2006**, *133*, 81–87.
28. Scott-Dupree, C.D.; Conroy, L.; Harris, C.R. Impact of currently used or potentially useful insecticides for canola agroecosystems on *Bombus impatiens* (Hymenoptera: Apidae), *Megachile rotundata* (Hymenoptera: Megachilidae), and *Osmia lignaria* (Hymenoptera: Megachilidae). *J. Econ. Entomol.* **2009**, *102*, 177–182.
29. Malone, L.A.; Scott-Dupree, C.D.; Todd, J.H.; Ramankutty, P. No sub-lethal toxicity to bumblebees, *Bombus terrestris*, exposed to Bt-corn pollen, captan and novaluron. *N. Z. J. Crop Hortic. Sci.* **2007**, *35*, 435–439.
30. Survilene, E.; Raudonis, L.; Jankauskiene, J. Investigation of pesticides effect on pollination of bumblebees in greenhouse tomatoes. *Sci. Works Lith. Inst. Hortic. Lith. Univ. Agric.* **2009**, *28*, 235–241.
31. Elston, C.; Thompson, H.M.; Walters, K.F.A. Sub-lethal effects of thiamethoxam, a neonicotinoid pesticide, and propiconazole, a DMI fungicide, on colony initiation in bumblebee (*Bombus terrestris*) micro-colonies. *Apidologie* **2013**, *44*, 563–574.
32. Gianessi, L.; Reigner, N. The value of fungicides in U.S. crop production. Available online: <http://croplife.intraspin.com/Pesticides/paper.asp?id=442> (accessed on 5 March 2015)
33. Van Scoy, A.R.; Tjeerdema, R.S. Environmental fate and Toxicology of chlorothalonil. *Rev. Environ. Contam. Toxicol.* **2014**, *232*, 89–105.
34. Koch, P.L.; Stier, J.C.; Senseman, S.A.; Sobek, S.; Kerns, J.P. Modification of a commercially-available ELISA kit to determine chlorothalonil and iprodione concentration on golf course turfgrass. *Crop Prot.* **2013**, *54*, 35–42.
35. De Ibarra, N.H.; Philippides, A.; Riabinina, O.; Collett, T.S. Preferred viewing directions of bumblebees (*Bombus terrestris* L.) when learning and approaching their nest site. *J. Exp. Biol.* **2009**, *212*, 3193–3204.
36. SAS Institute Inc. *JMP Pro 10*; SAS Institute Inc.: Cary, NC, USA, 2007.
37. Everich, R.; Schiller, C.; Whitehead, J.; Beavers, M.; Barrett, K. Effects of Captan on *Apis mellifera* brood development under field conditions in California almond orchards. *J. Econ. Entomol.* **2009**, *102*, 20–29.
38. Heinrich, B. *Bumblebee Economics*; Harvard University Press: Cambridge, MA, USA, 2004.

39. Pomeroy, N.; Plowright, R.C. The relation between worker numbers and the production of males and queens in the bumble bee *Bombus perplexus*. *Can. J. Zool.* **1982**, *60*, 954–957.
40. Mattila, H.R.; Rios, D.; Walker-Sperling, V.E.; Roeselers, G.; Newton, I.L.G. Characterization of the active microbiotas associated with honey bees reveals healthier and broader communities when colonies are genetically diverse. *PLoS ONE* **2012**, *7*, e32962.
41. Gilliam, M. Identification and roles of non-pathogenic microflora associated with honey bees1. *FEMS Microbiol. Lett.* **1997**, *155*, 1–10.
42. Kariluoto, S.; Aittamaa, M.; Korhola, M.; Salovaara, H.; Vahteristo, L.; Piironen, V. Effects of yeasts and bacteria on the levels of folates in rye sourdoughs. *Int. J. Food Microbiol.* **2006**, *106*, 137–143.
43. Yoder, J.A.; Jajack, A.J.; Rosselot, A.E.; Smith, T.J.; Yerke, M.C.; Sammataro, D. Fungicide contamination reduces beneficial fungi in bee bread based on an area-wide field study in honey bee, *Apis mellifera*, colonies. *J. Toxicol. Environ. Health A* **2013**, *76*, 587–600.

© 2015 by the authors; licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution license (<http://creativecommons.org/licenses/by/4.0/>).